

# **The development of novel optical screening tests for the presumptive identification of New Psychoactive Substances (NPS) in seized illicit materials**

**Morgan Philp**

A thesis submitted for the Degree of Doctor of Philosophy  
(Science)

University of Technology Sydney

July 2018

*This page intentionally left blank*

## Certificate of original authorship

I certify that the work in this thesis has not previously been submitted for a degree nor has it been submitted as part of requirements for a degree except as part of the collaborative doctoral degree and/or fully acknowledged within the text.

I also certify that the thesis has been written by me. Any help that I have received in my research work and the preparation of the thesis itself has been acknowledged. In addition, I certify that all information sources and literature used are indicated in the thesis.

This research is supported by an Australian Government Research Training Program Scholarship.

Production Note:  
Signature removed prior to publication.

---

Morgan Philp

25/07/2018

---

Date

# Acknowledgements

This thesis could not have been completed without the support I received from many others in my work and personal life and I would like to take this opportunity to extend my gratitude to all those people here.

To my supervisor Shanlin, your calm, caring and generous nature always kept me level-headed. I value the trust you placed in me throughout my PhD from project direction to representing our research group overseas. Your continued encouragement and support did not go unnoticed and I have nothing but good memories of my time as your student and I will be forever grateful that I chose your Honours project in 2012 all those years ago.

To my co-supervisor Ron, the labs at UTS just would not be the same without you in them. Thank you for your ongoing encouragement and passion for seeing through all projects successfully. For a busy man, you always found time to check on my progress and share your chemical knowledge with me. I am very grateful to be one of the lucky ones who have you on their supervisory panel.

A special thank you to Shanlin's toxicology research group (past and present members) for your support and for providing some normality to this PhD lifestyle. Shout-out to Dan, my fellow NPS researcher who I have shared this journey with. We have come a long way since our first ANZFSS conference in Adelaide and you have made my time here considerably more enjoyable. I look forward to seeing your conference Vans once more.

To the number of desk buddies I had during my time at UTS, you have honestly all kept me sane throughout this process. Kia, Tanya and Mel (Friday night Dream Team) during the early days, and Alicia, Rolanda and Mac in the later half. Thank you for putting a smile on my face and breaking up my long hours in the lab.

To Scott, your ability to go from the comic of the group to offering trusted and meaningful advice whenever needed is incredible. These are your strengths and what makes you the great, young academic you have become. Thank you for sharing your PhD woes and providing a glimmer of hope for post-PhD normalcy.

I would like to thank my family and friends for putting up with me throughout this PhD and especially for their incredibly encouraging words prior to submission. I truly appreciate you all. To my mum, Terri, thank you for allowing our phone calls to be hijacked by me talking about polymers and for reading through thesis chapters in your spare time.

To Christian, there are so many things I am grateful of, but most importantly is your dedication to standing with me while I went on this PhD journey. Thank you for being there after late nights, long hours and failed experiments. Your genuine praise and recognition at each turn did not go unnoticed.

“[This PhD research] was the best of times, it was the worst of times...”

- Charles Dickens, Tale of Two Cities, 1859

# Table of Contents

CERTIFICATE OF ORIGINAL AUTHORSHIP.....	III
ACKNOWLEDGEMENTS.....	IV
LIST OF FIGURES.....	XIV
LIST OF TABLES.....	XXIV
ABBREVIATIONS.....	XXVII
ABSTRACT.....	31
CHAPTER 1: INTRODUCTION.....	35
1.1 NEW PSYCHOACTIVE SUBSTANCES .....	37
1.1.1 Current situation.....	37
1.1.2 Synthetic cathinones.....	37
1.2 ILLICIT DRUG IDENTIFICATION TECHNIQUES .....	41
1.3 PHYSICAL EXAMINATION .....	43
1.4 CHEMICAL COLOUR TESTS .....	44
1.4.1 Historical overview .....	45
1.4.2 Chemistry of common colour tests .....	46
1.4.3 Drug class selective.....	52
1.4.4 Drug test kits.....	57
1.4.5 Harm reduction.....	58
1.4.6 New Psychoactive Substances .....	59
1.4.7 Limitations .....	61
1.4.8 Spot test regulations.....	62
1.4.9 Method validation guidelines .....	63

---

1.4.10	<i>Advances in colour testing</i> .....	65
1.4.11	<i>Future of presumptive colour testing</i> .....	69
1.5	MICROCRYSTALLINE TESTS .....	70
1.6	THIN LAYER CHROMATOGRAPHY (TLC) .....	71
1.6.1	<i>History and use</i> .....	71
1.6.2	<i>TLC in drug screening</i> .....	72
1.6.3	<i>Commercially available TLC</i> .....	73
1.6.4	<i>Current developments in TLC</i> .....	73
1.7	PORTABLE SPECTROSCOPIC ANALYSIS .....	74
1.7.1	<i>Fourier-Transform Infrared (FT-IR) Spectroscopy</i> .....	75
1.7.2	<i>Raman spectroscopy</i> .....	76
1.8	OPTICAL DETECTION METHODS .....	78
1.9	MOLECULAR RECOGNITION .....	79
1.9.1	<i>Selective chemical reactions</i> .....	79
1.9.2	<i>Molecularly Imprinted Polymers (MIPs)</i> .....	85
1.10	VISUAL SIGNALS .....	86
1.10.1	<i>Types of luminescence</i> .....	86
1.10.2	<i>Förster Resonance Energy Transfer (FRET)</i> .....	87
1.10.3	<i>Fluorescence quenching</i> .....	88
1.10.4	<i>Luminescence reporters</i> .....	88
1.11	MOLECULARLY IMPRINTED PHOTONIC HYDROGELS (MIPHs) .....	92
1.12	SIGNIFICANCE OF PRESUMPTIVE SCREENING TESTS .....	93
1.13	REFERENCES .....	94

<b>CHAPTER 2: SYNTHESIS OF CATHINONE ANALOGUES .....</b>	<b>113</b>
2.1 INTRODUCTION .....	113
2.2 MATERIALS AND METHODS .....	114
2.2.1 Chemicals.....	114
2.2.2 Melting point determination .....	114
2.2.3 Nuclear Magnetic Resonance (NMR) spectroscopy analysis.....	115
2.2.4 Gas Chromatography-Mass Spectrometry (GC-MS) analysis .....	115
2.2.5 Fourier Transform Infrared (FTIR) spectroscopy analysis .....	116
2.2.6 Ultraviolet-Visible spectroscopy analysis.....	116
2.2.7 Thin Layer Chromatography (TLC) .....	116
2.2.8 4-Methylmethcathinone (4-MMC, 3).....	117
2.2.9 4-Fluoromethcathinone (4-FMC, 6) .....	118
2.2.10 1-(1,3-benzodioxol-5-yl)-2-(methylamino)propan-1-one (methyllone, 11) .....	120
2.2.11 Methcathinone (MCAT, 13) .....	122
2.2.12 4-Methylethcathinone (4-MEC, 14) .....	123
2.2.13 4-Methylpyrrolidinopropiophenone (4-MPPP, 15) .....	124
2.2.14 3,4-Methylenedioxypropyvalerone (MDPV, 19) .....	125
2.2.15 1-(1,3-benzodioxol-5-yl)-2-(methylamino)butan-1-one (butyllone, 23) .....	127
2.2.16 1-(1,3-benzodioxol-5-yl)-2-(methylamino)pentan-1-one (pentyllone, 24) .....	129
2.2.17 1-(4-methylphenyl)-2-(pyrrolidin-1-yl)pentan-1-one (pyrovalerone, 29) .....	130
2.2.18 $\alpha$ -Pyrrolidinopentiophenone ( $\alpha$ -PVP, 34).....	132
2.2.19 4-Ethylmethcathinone (4-EMC, 39) .....	134
2.2.20 1-(4-methoxyphenyl)-2-(methylamino)pentan-1-one (MOMV, 44) .....	136
2.2.21 1-(2H-1,3-benzodioxol-5-yl)-2-(ethylamino)propan-1-one (ethylone, 45).....	138
2.2.22 2-(methylamino)-1-(naphthalen-1-yl)propan-1-one ( $\alpha$ -naphth, 50) .....	140



2.2.23	1-(2H-1,3-benzodioxol-5-yl)-2-(pyrrolidin-1-yl)butan-1-one (MDPBP, 51).....	142
2.2.24	1-(4-methylphenyl)-2-(pyrrolidin-1-yl)butan-1-one (4-MPBP, 55) .....	143
2.2.25	1-(4-hydroxyphenyl)-2-(methylamino)propan-1-one (4-HMC, 60) .....	146
2.2.26	2-(methylamino)-1-(naphthalen-2-yl)pentan-1-one ( $\beta$ -naphyrone, 65) .....	147
2.2.27	2-Amino-1-phenylpropan-1-one (CAT, 70) .....	149
2.3	RESULTS AND DISCUSSION .....	150
2.3.1	Reaction summary .....	150
2.3.2	Yield and melting point .....	153
2.3.1	Characterisation .....	153
2.3.2	Unsuccessful syntheses .....	163
2.4	CONCLUSION .....	163
2.5	REFERENCES .....	164

### CHAPTER 3: CHEMICAL COLOUR TESTS FOR THE COLORIMETRIC DETECTION OF SYNTHETIC CATHINONES 170

SUMMARY .....	170
3.1 INTRODUCTION .....	172
3.2 MATERIALS AND METHODS .....	174
3.2.1 Chemicals .....	174
3.2.2 Reference materials .....	174
3.2.3 Preparation of working solutions .....	175
3.2.4 Apparatus and instrumentation .....	175
3.2.5 Colour test method development .....	176
3.2.6 Spectroscopic analysis .....	178
3.2.7 Method validation .....	179
3.3 RESULTS AND DISCUSSION .....	181

## Table of Contents

---

3.3.1	<i>Colour test method development</i> .....	181
3.3.2	<i>General recommended procedure</i> .....	185
3.3.3	<i>Spectroscopic analysis</i> .....	186
3.3.4	<i>Method validation</i> .....	190
3.4	CONCLUSION .....	200
3.5	REFERENCES .....	200
 <b>CHAPTER 4: INVESTIGATION INTO A CHEMICAL COLOUR TEST DEVICE</b> .....		<b>206</b>
4.1	INTRODUCTION .....	206
4.2	MATERIALS AND METHODS .....	207
4.2.1	<i>Chemicals</i> .....	207
4.2.2	<i>Reference material</i> .....	207
4.2.3	<i>Apparatus and materials</i> .....	207
4.2.4	<i>Investigation of friction and heat generating mechanisms</i> .....	208
4.2.5	<i>Investigation of catalysts and heat activators</i> .....	210
4.2.6	<i>Combination of friction/heating mechanism and catalyst</i> .....	211
4.2.7	<i>Preliminary investigation of paper test strips (Experiment 9)</i> .....	211
4.2.8	<i>Investigation into test simplicity and adaptability</i> .....	212
4.2.9	<i>Creation of a simple colour test method</i> .....	213
4.3	RESULTS .....	214
4.3.1	<i>Investigation of friction and heat generating mechanisms</i> .....	214
4.3.2	<i>Investigation of catalysts and heat activators</i> .....	217
4.3.1	<i>Combination of friction/heating mechanism and catalyst</i> .....	219
4.3.2	<i>Preliminary investigation of paper test strips (Experiment 9)</i> .....	220
4.3.1	<i>Investigation into test simplicity and adaptability</i> .....	221

---

4.3.1	<i>Creation of a simple colour test method.....</i>	222
4.4	DISCUSSION .....	223
4.5	CONCLUSION .....	229
4.6	REFERENCES .....	230
 <b>CHAPTER 5: CHEMICAL REACTIONS OF SYNTHETIC CATHINONES: A POTENTIAL RECOGNITION ELEMENT? .....</b>		<b>234</b>
5.1	INTRODUCTION .....	234
5.2	MATERIALS AND METHODS.....	235
5.2.1	<i>Chemicals .....</i>	235
5.2.2	<i>Reference material.....</i>	235
5.2.3	<i>Instrumentation .....</i>	235
5.2.4	<i>Ultraviolet-Visible spectroscopy analysis.....</i>	236
5.2.5	<i>Thin Layer Chromatography (TLC).....</i>	236
5.2.6	<i>Gas Chromatography-Mass Spectrometry (GC-MS) analysis.....</i>	236
5.2.7	<i>Preparation of cathinone stock solutions.....</i>	237
5.2.8	<i>Imine (and enamine) formation .....</i>	237
5.2.9	<i>Hydrazone formation .....</i>	238
5.2.10	<i>Semicarbazone formation.....</i>	239
5.2.11	<i>Oxime formation .....</i>	240
5.2.12	<i>Reduction reactions.....</i>	242
5.2.13	<i>Antioxidant activity .....</i>	242
5.2.14	<i>Cathinone stability studies .....</i>	244
5.3	RESULTS.....	246
5.3.1	<i>Imine (and enamine) formation .....</i>	246
5.3.2	<i>Hydrazone formation .....</i>	246

5.3.3	<i>Semicarbazone formation</i> .....	247
5.3.4	<i>Oxime formations</i> .....	247
5.3.5	<i>Reduction reactions</i> .....	249
5.3.6	<i>Antioxidant activity</i> .....	250
5.3.7	<i>Cathinone stability studies</i> .....	252
5.4	DISCUSSION .....	257
5.5	CONCLUSION .....	260
5.6	REFERENCES .....	260

## **CHAPTER 6: PREPARATION AND APPLICATION OF MOLECULARLY IMPRINTED POLYMERS (MIPS) 265**

6.1	INTRODUCTION .....	265
6.2	MATERIALS AND METHODS .....	266
6.2.1	<i>Chemicals</i> .....	266
6.2.2	<i>Reference material</i> .....	266
6.2.3	<i>Instrumentation</i> .....	266
6.2.4	<i>Ultraviolet-visible (UV-Vis) spectroscopy analysis</i> .....	266
6.2.5	<i>Gas Chromatography-Mass Spectrometry (GC-MS) analysis</i> .....	267
6.2.6	<i>Nuclear Magnetic Resonance (NMR) spectroscopy study</i> .....	267
6.2.7	<i>Preparation of Molecularly Imprinted Polymers (MIPs)</i> .....	267
6.2.8	<i>Cathinone imprinted polymers</i> .....	270
6.2.9	<i>Melamine-Urea-Formaldehyde (MUF) resins</i> .....	272
6.2.10	<i>Molecular interaction studies</i> .....	273
6.3	RESULTS AND DISCUSSION .....	274
6.3.1	<i>Preparation of MIPS</i> .....	274
6.3.2	<i>Cathinone imprinted polymers</i> .....	278

6.3.3	<i>MUF resins</i> .....	279
6.3.4	<i>Molecular interaction studies</i> .....	281
6.4	CONCLUSION .....	282
6.5	REFERENCES .....	283
 <b>CHAPTER 7: CONCLUSIONS AND FUTURE WORK</b> .....		<b>287</b>
7.1	CONCLUDING REMARKS .....	287
7.2	FUTURE WORK .....	289
 <b>APPENDICES</b> .....		<b>291</b>

## List of Figures

Figure 1-1. Fresh khat leaves (from Palmer, 2010)[11] .....	38
Figure 1-2. Generic structure of synthetic cathinone analogues showing the substitution possibilities that led to formation of a wide range of compounds .....	39
Figure 1-3. Classification of 15 common colour test reagents categorised by the pH of the test solution and the type of coloured product formed. ....	48
Figure 1-4. Chemical reactions of common colour tests that are selective toward a range of drug classes and used in general screening methods. Marquis reagent (a); Liebermann's reagent (b); Mandelin's reagent (c); and Froehde's reagent (d).....	50
Figure 1-5. Chemical reactions of common colour spot tests that are selective toward certain functional groups. Simon's reagent for secondary amines (a); Zimmerman reagent for $\beta$ -amino ketones (b); and ferric chloride reagent for phenols (c). ....	51
Figure 1-6. Chemical reactions of common colour spot tests that are selective toward a drug class. Duquenois-Levine reagent for cannabinoids (a); Mecke reagent for opium alkaloids (b); Scott's reagent for cocaine (c); Dille-Koppanyi reagent for barbiturates (d). ....	54
Figure 1-7. Chemical reactions of common colour spot tests that are selective toward a drug class. Chen-Kao reagent for ephedrine/norephedrine (a); Ehrlich's reagent for ergot alkaloids (and LSD) (b); Fast Blue BB reagent for cannabinoids (c); and Zwikker reagent for barbiturates (d). ....	55
Figure 1-8. Screening methods performed by laboratories taking part in the UNODC's International Collaborative Exercises (ICE) study of seized materials over three separate studies from 2014-2016.....	56
Figure 1-9. Reaction equation for the formation of an imine from a ketone .....	79
Figure 1-10. Reaction equation for the formation of a hydrazone derivative from a ketone ....	81
Figure 1-11. Reaction equation for the formation of an oxime from a ketone .....	81

Figure 1-12. Reaction equation for the formation of a semicarbazone from a ketone .....	82
Figure 1-13. Schematic of the preparation process of molecularly imprinted polymers.....	85
Figure 1-14. Simplified energy level diagram illustrating FRET between an excited donor fluorophore (D*) and an acceptor molecule (A). Horizontal arrows demonstrate coupled transitions .....	87
Figure 1-15. Absorption and emission spectra of two fluorophores exhibiting FRET. The shaded area demonstrates the overlap of donor emission and acceptor absorbance .....	88
Figure 1-16. Structure of organic dyes: fluorescein (A) and BODIPY parent fluorophore (B) ....	89
Figure 1-17. Schematic representation of the preparation of molecularly imprinted photonic hydrogels (MIPHs).....	93
Figure 2-1. Synthesis of 4-methylmethcathinone HCl ( <b>3</b> ) via a two step reaction sequence: $\alpha$ -bromination of <b>1</b> followed by nucleophilic substitution of <b>2</b> with methylamine. The final product salt was obtained following extraction and HCl work-up.....	117
Figure 2-2. Synthesis of 4-fluoromethcathinone HCl ( <b>6</b> ) via a two step reaction sequence: $\alpha$ -bromination of <b>4</b> followed by nucleophilic substitution of <b>5</b> with methylamine. The final product salt was obtained following extraction and HCl work-up.....	118
Figure 2-3. Synthesis of methylone HCl ( <b>11</b> ) via a four step reaction sequence: Grignard reaction of <b>7</b> , oxidation of <b>8</b> , $\alpha$ -bromination of <b>9</b> , and nucleophilic substitution of <b>10</b> with methylamine. The final product salt was obtained following extraction and HCl work-up. ....	120
Figure 2-4. Synthesis of methcathinone HCl ( <b>13</b> ) via oxidation of ephedrine HCl ( <b>12</b> ). The final product salt was obtained following extraction and HCl work-up. ....	122
Figure 2-5. Synthesis of 4-methylethcathinone HCl ( <b>14</b> ) via a two step reaction sequence: $\alpha$ -bromination of <b>1</b> followed by nucleophilic substitution of <b>2</b> with ethylamine. The final product salt was obtained following extraction and HCl work-up.....	123
Figure 2-6. Synthesis of 4-methylpyrrolidinopropiophenone HCl ( <b>15</b> ) via a two step reaction sequence: $\alpha$ -bromination of <b>1</b> followed by nucleophilic substitution of <b>2</b> with pyrrolidine. The final product salt was obtained following extraction and HCl work-up. ....	124

Figure 2-7. Synthesis of MDPV HCl (**19**) via a four step reaction sequence: Grignard reaction of piperonal (**7**), oxidation of **16**,  $\alpha$ -bromination of **17** and nucleophilic substitution of **18** with pyrrolidine. The final product salt was obtained following extraction and HCl work-up. .... 125

Figure 2-8. Synthesis of butylone HCl (**23**) via a four step reaction sequence: Grignard reaction of piperonal (**7**), oxidation of **20**,  $\alpha$ -bromination of **21** and nucleophilic substitution of **22** with methylamine. The final product salt was obtained following extraction and HCl work-up..... 127

Figure 2-9. Synthesis of pentylone HCl (**24**) via a four step reaction sequence: Grignard reaction of piperonal (**7**), oxidation of **16**,  $\alpha$ -bromination of **17** and nucleophilic substitution of **18** with methylamine. The final product salt was obtained following extraction and HCl work-up..... 129

Figure 2-10. Synthesis of pyrovalerone HCl (**29**) via a four step reaction sequence: Grignard reaction of *p*-tolualdehyde (**25**), oxidation of **26**,  $\alpha$ -bromination of **27** and nucleophilic substitution of **28** with pyrrolidine. The final product salt was obtained following extraction and HCl work-up. .... 130

Figure 2-11. Synthesis of  $\alpha$ -PVP HCl (**34**) via a four step reaction sequence: Grignard reaction of benzonitrile (**30**), oxidation of **31**,  $\alpha$ -bromination of **32**, and nucleophilic substitution of **33** with pyrrolidine. The final product salt was obtained following extraction and HCl work-up. .... 132

Figure 2-12. Synthesis of 4-EMC HCl (**39**) via a four step reaction sequence: Grignard reaction of *p*-ethylbenzaldehyde (**35**), oxidation of **36**,  $\alpha$ -bromination of **37**, and nucleophilic substitution of **38** with methylamine. The final product salt was obtained following extraction and HCl work-up. .... 134

Figure 2-13. Synthesis of MOMV HCl (**44**) via a four step reaction sequence: Grignard reaction of *p*-methoxybenzaldehyde (**40**), oxidation of **41**,  $\alpha$ -bromination of **42**, followed by amination of **43** with methylamine. The final product salt was obtained following extraction and HCl work-up. .... 136

Figure 2-14. Synthesis of ethylone HCl (**45**) via a four step reaction sequence: Grignard reaction of piperonal (**7**), oxidation of **8**,  $\alpha$ -bromination of **9**, followed by amination of **10** with ethylamine. The final product salt was obtained following extraction and HCl work-up..... 138



- Figure 2-15. Synthesis of  $\alpha$ -naphth HCl (**50**) via a four step reaction sequence: Grignard reaction of  $\alpha$ -naphthaldehyde (**46**), oxidation of **47**,  $\alpha$ -bromination of **48**, amination of **49** with methylamine. The final product salt was obtained following extraction and HCl work-up. .... 140
- Figure 2-16. Synthesis of MDPBP HCl (**51**) via a four step reaction sequence: Grignard reaction of piperonal (**7**), oxidation of **20**,  $\alpha$ -bromination of **21**, followed by amination of **22** with pyrrolidine. The final product salt was obtained following extraction and HCl work-up. .... 142
- Figure 2-17. Synthesis of 4-MPBP HCl (**55**) via a four step reaction sequence: Grignard reaction of *p*-tolualdehyde (**25**), oxidation of **52**,  $\alpha$ -bromination of **53** and nucleophilic substitution of **54** with pyrrolidine. The final product salt was obtained following extraction and HCl work-up. 143
- Figure 2-18. Theoretical synthesis of 4-HMC HCl (**60**) via a four step reaction sequence: Grignard reaction of *p*-hydroxybenzaldehyde (**56**), oxidation of **57**,  $\alpha$ -bromination of **58** and nucleophilic substitution of **59** with methylamine. .... 146
- Figure 2-19. Synthesis of  $\beta$ -naphyrone HCl (**65**) via a four step reaction sequence: Grignard reaction of 2-naphthaldehyde (**61**), oxidation of **62**,  $\alpha$ -bromination of **63** and nucleophilic substitution of **64** with pyrrolidine. The final product salt was obtained following extraction and HCl work-up. .... 147
- Figure 2-20. Theoretical synthesis of cathinone HCl (**69**) via a four step reaction sequence: Grignard reaction of benzonitrile (**30**), oxidation of **66**,  $\alpha$ -bromination of **67** and amination of **68** either directly, or via a phthalimide derivative (**70**). .... 149
- Figure 2-21. Overall reaction scheme for the preparation of synthetic cathinones. *Reactions: (i) and (ii)* Grignard reaction followed by oxidation; *(iii)* Bromination; *(iv)* Methamination; *(v)* Ethamination; *(vi)* Amination with pyrrolidine; *(vii)* Oxidation. .... 151
- Figure 2-22. Reaction mechanism pathway for the synthesis of a general cathinone analogue. .... 152
- Figure 2-23. Total ion chromatograms showing significant peak tailing potentially due to unresolved artefacts formed during GC-MS analysis. A: MCAT (**13**); B: 4-EMC (**39**). .... 156

Figure 2-24. Alpha-cleavage reaction of synthetic cathinones to produce the major immonium cation fragments. Alkyl chain length and N-substituent of the cathinone determines the base peak $m/z$ . .....	157
Figure 2-25. EI mass spectrum of butylone HCl ( <b>23</b> ) showing major fragments ions at $m/z$ 72, 121, 149 and 192 due to $\alpha$ -cleavage fragmentation pathways. ....	157
Figure 2-26. Annotated ATR-FTIR spectrum of ethylone HCl ( <b>45</b> ).....	158
Figure 2-27. Expected proton coupling ( $^1\text{H}$ - $^1\text{H}$ ) on monosubstituted, <i>p</i> -disubstituted and trisubstituted benzene rings.....	160
Figure 2-28. Fine structure observed downfield in the $^1\text{H}$ NMR spectrum of 4-FMC ( <b>6</b> ) as a result of heteronuclear coupling between $^1\text{H}$ and $^{19}\text{F}$ . Extra peaks observed in the spectrum are due to the added complexity of coupling between magnetically nonequivalent $\text{H}_\text{A}$ and $\text{H}_\text{A}'$ ( $\text{H}_\text{B}$ and $\text{H}_\text{B}'$ ) .....	160
Figure 2-29. Upfield of the pentylone ( <b>24</b> ) $^1\text{H}$ NMR spectrum showing methylene proton splitting patterns .....	161
Figure 2-30. Upfield of $^1\text{H}$ NMR spectrum of $\alpha$ -PVP ( <b>34</b> ) showing signals due to pyrrolidine ring protons. ....	162
Figure 3-1. General chemical structure of synthetic cathinone substances. $\text{R}_1$ and $\text{R}_2$ can exist as hydrogen, an alkyl moiety, or cyclic structure; $\text{R}_3$ can exist as hydrogen or any alkyl group; and $\text{R}_4$ can exist as hydrogen or a combination of various moieties such as alkyl, alkoxy, alkylendioxy, haloalkyl, or halide. ....	172
Figure 3-2. Preliminary copper-neocuproine test results. Method performed on control reagent blank ( <i>a</i> ) and aqueous 4-MMC HCl solution ( <i>b</i> ). Coloured solutions were pipetted from beakers into white well-plates after heating to improve the colour contrast.....	181
Figure 3-3. Reagent concentration optimization study results. Colour testing performed using decreasing neocuproine concentrations (1-5) and decreasing acetate concentrations ( <i>a-e</i> ) with $\text{Cu(II)}$ concentration kept at $1.25 \times 10^{-2}$ M for control reagent blank ( <i>left</i> ) and aqueous 4-MMC HCl ( <i>right</i> ). The well at 5b in the control blank and 4-MMC HCl charged plate was chosen to have	

the optimal concentrations for testing: $1.25 \times 10^{-2}$ M Cu(II); $5.12 \times 10^{-3}$ M neocuproine; and 2.00 M acetate buffer. ....	183
Figure 3-4. Final copper-neocuproine test results. Method performed on control blank (1) and solid 4-MMC HCl (2) at two different stages: before heating (a) and after heating (b). ....	185
Figure 3-5. Overlaid Ultraviolet-Visible absorbance spectra of colour test results. Cu(II)-neocuproine colour test control reagent blank (1) and Cu(II)-neocuproine colour test on 4-MMC HCl (2). Inset: Zoomed region of the absorption band centered at 453 nm. ....	187
Figure 3-6. The residue/crystalline material remaining in the well-plate 48 hours after testing of control blank (a) and 4-MMC HCl sample (b). The combined crystalline materials were washed with diethyl ether and cold water; filtered through a cotton-plugged pipette; and collected by redissolving in organic solvent (c). ....	187
Figure 3-7. $^1\text{H}$ -NMR spectrum of extracted yellow-orange coloured product, $\text{Cu(I)(neocuproine)}_2$ , recorded in deuterated chloroform. Downfield (A) and upfield spectral regions (B) are defined. Proton environments have been assigned, with labeled peaks corresponding to the protons on the complex structure. Integration values are provided in brackets for each relevant chemical shift. ....	188
Figure 3-8. Reaction equation for the formation of the yellow-orange coloured complex. The copper metal center in $\text{Cu(II)(neocuproine)}_2$ is reduced in the presence of a reductant to form $\text{Cu(I)(neocuproine)}_2$ (A). 3-Dimensional molecular structures of $\text{Cu(II)(neocuproine)}_2$ (drawn without potential acetate or water ligands) and $\text{Cu(I)(neocuproine)}_2$ are proposed to demonstrate the difference in geometry and arrangement of the neocuproine chelating ligands (B). ....	189
Figure 3-9. Limit of detection method validation test results for 4-MMC HCl using the proposed neocuproine colour test method. Amounts of 4-MMC HCl subjected to testing are 0, 1, 2, 3, 4, 5, 6, 10, 20, 30, 40, 50 $\mu\text{g}$ (a-l). ....	198
Figure 4-1. Alternative heating devices used in Experiment 4 in replacement of the AC powered hot plate. A) USB-powered mug warmer and B) rechargeable electric cigarette lighter. ....	209
Figure 4-2. Schematic of the paper test strip used in preliminary work on a paper-based device .....	212

Figure 4-3. Test results using a combination of PE bag and glass beads (Exp 1-2) performed with A) reagents only as a control reagent blank and B) 4-MMC as a positive control .....	214
Figure 4-4. Test results using a combination of PE bag and microbeads performed with addition of 4-MMC after 5 min using A) PE pellets (Exp 3-1), B) glass beads (Exp 3-2) and C) PVAc solid resins (Exp 3-3) .....	215
Figure 4-5. Result of colour test (Exp 4-1) performed on 4-MMC as a positive control ( <i>left</i> ) and control reagent blank ( <i>right</i> ) using USB-powered mug warmer after 10 minutes of heating..	216
Figure 4-6. The use of the electronic cigarette lighter in colour test methods (Exp 4-2). A) The coil heating element on which the glass vial was intermittently placed and B) result of colour test performed on 4-MMC as positive control .....	217
Figure 4-7. Test results using small glass vial and surrounding exothermic reaction (Exp 4-3). A) control reagent blank after 8 min, B) 4-MMC positive control after 8 min, and C) 4-MMC after 1 h .....	217
Figure 4-8. Results of the most optimal combinations of heat activator and friction mechanism after 5 min (Exp 7). A) silica gel and glass beads, B) KF and glass beads, and C) silica gel and PE pellets .....	220
Figure 4-9. Colour test result after 4 minutes by combining the use of a catalyst with an external exothermic reaction as a simple heating mechanism (Exp 8). A) control blank with KCl, B) 4-MMC with KCl, C) control blank without KCl, D) 4-MMC without KCl .....	220
Figure 4-10. Result of preliminary paper test strip method (Exp 9) performed on a blank control ( <i>left</i> ) and 4-MMC ( <i>right</i> ). .....	221
Figure 4-11. Colour test result employing chloroform as an organic extraction solvent to concentrate the coloured product performed in A) a semi-micro test tube and B) a sealed Pasteur pipette .....	222
Figure 4-12. Results of limit of detection (LOD) study performed on 4-MMC after 2 minutes ( <i>top</i> ) and 5 minutes ( <i>bottom</i> ). Amounts between 0.1 µg and 0.8 µg are not shown here due to no colour change being observed. ....	223

Figure 4-13. Colour test device design employing glass beads sandwiched between two rotating glass discs .....	224
Figure 4-14. Colour test device design employing glass beads sandwiched between one concave and one convex shaped glass discs that rotate to generate friction .....	225
Figure 4-15. Colour test device design that employs a polyethylene pouch that contains a glass ampoule with organic solvent and colour test reagents .....	227
Figure 4-16. Colour test device design employing a collection swab that screws into a sealed tube containing the reagent mix and dichloromethane .....	228
Figure 4-17. Colour test device accessory that allows the coloured portion of the tapered tube to be recorded using a portable colour digitiser or smart phone application.....	229
Figure 5-1. Synthetic preparation of an imine from a synthetic cathinone and a primary amine .....	237
Figure 5-2. Synthetic preparation of 2,4-dinitrophenylhydrazone derivative from a synthetic cathinone and 2,4-dinitrophenylhydrazine (DNP) .....	238
Figure 5-3. Synthetic preparation of a semicarbazone derivative from a synthetic cathinone and semicarbazide hydrochloride.....	239
Figure 5-4. Synthetic preparation of an oxime derivative from a synthetic cathinone and hydroxylamine .....	240
Figure 5-5. The synthetic preparation of secondary alcohol of synthetic cathinones via a reduction reaction .....	242
Figure 5-6. Representative TIC of enamine formation method 2 product mixture using 4-MMC as starting material. ....	246
Figure 5-7. GC-MS analysis of the product of <i>oximation method 2</i> performed on 4-MMC. A) Representative TIC showing two major components are unreacted 4-MMC and the 4-MMC oxime derivative, B) Mass spectrum obtained for the peak at 6.725 min in the TIC trace .....	248

Figure 5-8. Developed and iodine stained TLC plates used in reduction reaction monitoring of A) pentylone, B) butylone, and C) pyrovalerone showing formation of new compounds.....	249
Figure 5-9. TIC trace of MDPV reduction product mixture showing the split peak observed in all pyrrolidine containing analogues .....	250
Figure 5-10. The absorbance of control blank, ascorbic acid, glucose and 10 cathinone analogues at 450 nm following the CUPRAC assay.....	251
Figure 5-11. The absorbance of control blank, ascorbic acid, glucose and 7 cathinone analogues at 695 nm following a total antioxidant capacity measurement .....	252
Figure 5-12. UV spectra of ‘propiophenone’ analogues stored in pH 10 buffer at room temperature over a one month period. 4-FMC and MCAT showed significant degradation after one week storage. ....	254
Figure 5-13. UV spectra of A) methylone HCl and B) pyrovalerone HCl stored in pH 10 buffer at room temperature over a one month period .....	254
Figure 5-14. TIC of extracted sample of 4-MEC stored in pH 10 buffer after 28 days showing all 4-MEC had degraded to 4-methylbenzaldehyde.....	256
Figure 5-15. TIC of extracted sample of $\alpha$ -PVP stored in pH 10 buffer after 28 days showing small presence of degradation product, benzaldehyde .....	256
Figure 5-16. UV spectra of aqueous $\alpha$ -PVP HCl solution stored under UV light over a one month period .....	257
Figure 5-17. The electron donating substituents on the carbonyl group of a cathinone that effectively reduce the nucleophilicity of the C atom and reactivity of the carbonyl group .....	257
Figure 6-1. Bulk monolith polymers prepared from pre-polymerisation solutions of A) Experiment 1-1 (non-imprinted), B) Experiment 1-3 (imprinted), and C) Experiment 1-3 (imprinted and non-imprinted still in glass pipette) .....	275
Figure 6-2. UV-Vis spectra of resulting binding solutions after incubation of polymer 1-1 (80 mg) and vanillin solution (0.05 mM) for 12 h. ....	276

---

Figure 6-3. The polymerised solutions of ephedrine imprinted (left) and non-imprinted (right) polymers (pol-3).....	276
Figure 6-4. Difference in absorbance of ephedrine at 207 nm between non-imprinted polymer binding solutions and imprinted polymer binding solutions at different concentrations after 12 min .....	277
Figure 6-5. TIC from the analysis of methanol elution solvent of makeshift SPE tube from 4-MMC loading solution. Non-imprinted and imprinted polymers were examined.....	278
Figure 6-6. Preparation of MUF resins for selective adsorption: A) MUF gel mixture prior to incubation, B) imprinted and non-imprinted MUF resins after incubation, C) grinding MUF resins in mortar and pestle .....	279
Figure 6-7. UV spectrum of 4-MMC loading solution eluate after passing through a cartridge containing imprinted and non-imprinted MUF resins. The imprinted resin adsorbs more 4-MMC molecules, and decreases the 4-MMC concentration .....	280
Figure 6-8. NMR titration plot showing the change in chemical shift values of two key carbon signals as a function of molar equivalents of MAA added.....	281
Figure 6-9. NMR titration plot showing the change in chemical shift values of two key carbon signals as a function of molar equivalents of EGDMA are added.....	282

# List of Tables

Table 1-1. The names and chemical structures of common synthetic cathinones showing the location of the substituents to the benzene ring and amine group .....	39
Table 1-2. Colour test reagent compositions and targeted drugs for common colour spot tests used in illicit drug detection .....	46
Table 1-3. Commercial companies producing colour test kits for drugs of abuse and precursor material. ....	57
Table 1-4. Selection of companies producing commercial portable spectroscopy instruments for analysis of illicit substances in the field.....	77
Table 2-1. Percentage yields obtained for synthetic cathinone analogues and their respective ketone and bromo-ketone intermediates.....	154
Table 2-2. GC-MS data obtained for the isolated and characterised synthetic cathinone analogues.....	155
Table 2-3. Key FTIR absorption bands for the synthetic cathinone analogues .....	159
Table 2-4. UV-Visible absorption maxima of the synthetic cathinones in this study, recorded in deionised water .....	162
Table 3-1. Selectivity study results of proposed neocuproine colour test method with pure synthetic cathinone substances and in mixtures with other analytes .....	191
Table 3-2. Selectivity study results of proposed neocuproine colour test method with amphetamines, common precursor chemicals and other recreational drugs.....	194
Table 3-3. Selectivity study results of proposed neocuproine colour test method with a range of common adulterants, excipients and powdered substances.....	196
Table 4-1. Amounts of catalyst/activator employed in test trials .....	210



Table 4-2. Results of colour test device trials that employed friction generating mechanisms as an alternative to the hotplate.....	215
Table 4-3. Results of colour test device trials that employed heat generating methods alternative to the hotplate .....	216
Table 4-4. Results of colour test trials using catalysts and “heat activators” at room temperature .....	218
Table 4-5. Results of colour test trials using three different sized molecular sieves at room temperature.....	218
Table 4-6. Results of colour test trials combining a heat activator with a friction mechanism <sup>a</sup> .....	219
Table 4-7. Issues arising from the use of microbeads in a PE bag and possible solutions to these problems .....	223
Table 5-1. Storage solutions and environment conditions examined in stability studies performed on 10 synthetic cathinone analogues.....	244
Table 5-2. Results of the hydrazone derivative preparation performed on selected cathinones and intermediates containing carbonyl functional groups.....	247
Table 5-3. Results of oximation method 3 performed on 10 cathinone analogues .....	249
Table 5-4. Three cathinone analogue classifications based on substituents and structure .....	253
Table 5-5. Results of GC-MS analysis of pH 10 cathinone solutions after 28 days storage at room temperature.....	255
Table 6-1. Experiment parameters for methods adapted from the literature.....	268
Table 6-2. Binding studies performed on the ephedrine imprinted (and non-imprinted) polymer (pol-3).....	269
Table 6-3. Cathinone imprinted polymer preparation conditions.....	271
Table 6-4. Solution preparation for NMR interaction study of 4-MMC with monomer, MAA.	273

Table 6-5. Solution preparation for NMR interaction study of 4-MMC with cross-linker, EGDMA	
.....	274

# Abbreviations

$^{13}\text{C}$ -NMR	carbon nuclear magnetic resonance
$^1\text{H}$ -NMR	proton nuclear magnetic resonance
4-EMC	4-ethylmethcathinone
4-FMC	4-fluoromethcathinone
4-HMC	1-(4-hydroxyphenyl)-2-(methylamino)propan-1-one
4-MEC	4-methylethcathinone
4-MMC	4-methylmethcathinone
4-MPBP	1-(4-methylphenyl)-2-(pyrrolidin-1-yl)butan-1-one
4-MPPP	4-methylpyrrolidinopropiophenone
AC	alternating current
ACN	acetonitrile
amu	atomic mass unit
approx.	approximately
aq.	aqueous
atm	atmosphere
ATR	attenuated total reflectance
br.	broad
BuMgCl	butylmagnesium chloride
CAT	2-amino-1-phenylpropan-1-one
conc.	concentrated
d	doublet
DCM	dichloromethane
dd	doublet of doublet
dec.	decomposed
deform.	deformation
EI	electron ionisation
Et <sub>2</sub> O	diethyl ether
EthMgCl	ethylmagnesium chloride
FTIR	Fourier transform infrared
GC-MS	gas chromatography-mass spectrometry
h	hours
J	coupling constant

## Abbreviations

---

lit.	literature value
M	moles per litre
m	multiplet
m/z	mass to charge ratio
M+	molecular ion
MCAT	methcathinone
MDPBP	1-(2H-1,3-benzodioxol-5-yl)-2-(pyrrolidin-1-yl)butan-1-one
MDPBP	1-(2H-1,3-benzodioxol-5-yl)-2-(pyrrolidin-1-yl)butan-1-one
MDPV	3,4-methylenedioxypyrovalerone
MDPV	3,4-methylenedioxypyrovalerone
MeOH	methanol
min	minutes
MIP	molecularly imprinted polymer
MIPH	molecularly imprinted photonic hydrogels
MOMV	1-(4-methoxyphenyl)-2-(methylamino)pentan-1-one
mp	melting point
MW	molecular weight
NPS	new psychoactive substance
∅	diameter
oop	out of plane
PCC	pyridinium chlorochromate
PE	polyethylene
ppm	parts per million
ppt.	precipitate
PropMgCl	propylmagnesium chloride
PVAc	polyvinyl acetate
q	quartet
RBF	round bottom flask
Ref.	references
R <sub>f</sub>	retardation factor
RT	room temperature
s	singlet
SN <sub>2</sub>	one step nucleophilic substitution reaction
t	triplet
THF	tetrahydrofuran

TIC	total ion chromatogram
TLC	thin layer chromatography
TMS	tetramethylsilane
UCNP	upconversion nanoparticles
unk.	unknown
unsym.	unsymmetrical
UV-Vis	ultraviolet-visible
$\alpha$ -PVP	$\alpha$ -pyrrolidinopentiophenone
$\delta$	chemical shift
$\lambda$	wavelength
$\bar{\nu}$	wavenumber

*This page intentionally left blank*

# Abstract

The large and increasing number of illicit materials seized each year combined with the introduction of many new psychoactive substances (NPS) to the traditional drug market are concerning realities. The need for simple presumptive field tests able to accurately detect compounds such as synthetic cathinones with good sensitivity is apparent. Field testing is an important tool for law enforcement officers to obtain rapid feedback regarding an unknown substance while awaiting confirmatory analysis results often in a backlog.

Chemical reactions selective toward the synthetic cathinone molecular structure were investigated for their application as a receptor element in an optical screening test.

Oximations, hydrazone formations, semicarbazide formations and metal complexation were examined with a number of cathinone analogues. As an alternative to chemical reactions, molecularly imprinted polymers (MIPs) selective toward synthetic cathinone compounds were prepared, optimised and tested for their selective binding ability.

The synthetic cathinone class of compounds failed to show significant reactivity with the reagents selected under a range of experimental conditions. However, their ability to actively reduce the cupric ion of a copper-neocuproine complex to afford a yellow-orange coloured product was observed. This colour test was optimised, validated and later improved upon to increase useability in the field as an optical screening test. The MIPs showed potential in selectively recognising the presence of the target cathinone molecule through binding studies performed, however, before the polymer can be used as a receptor, further optimisation is required.

The three receptors investigated provided different degrees of success. The chemical colour test was successfully developed into a protocol for application in field testing of cathinones; the MIPs showed some potential for further investigation and application in a different protocol for cathinone detection; and the use of chemical reactions for tagging purposes was not achieved due to the inherently unreactive carbonyl group of the cathinone molecular structure. Nonetheless, this research provided significant and useful chemical analysis of the synthetic cathinone class of NPS, while raising awareness of current deficiencies in the presumptive identification of NPS.

*This page intentionally left blank*



# **Chapter 1: Introduction**

*This page intentionally left blank*